

Exome CNV Overlapping (ECO): an integrative copy number variation caller for exome sequencing

Peng Zhang, Hua Ling, Elizabeth Pugh, Kimberly Doheny

Center for Inherited Disease Research (CIDR), Johns Hopkins Genomics, Institute of Genetic Medicine, The Johns Hopkins School of Medicine

Introduction

Due to the uneven distribution of reads and the sparse nature of target regions for whole exome sequencing (WES) data, calling copy number variations (CNVs) has been a challenge. Most existing programs can only use read counts as inputs and calls often vary between programs. As part of the validation process, we found that some confirmed causal CNVs were called by multiple programs while others were not. In addition, each program often requires different input files and its output format often varies with different breakpoints for the CNV calls, which makes it difficult to compare and summarize results across programs.

We present here a practical pipeline that integrates multiple CNV calling programs and generates one combined VCF-like report with merged calls and annotations. It incorporated three prevalent CNV calling programs (ExomeDepth [Plagnol et al. 2012], CANOES [Backenroth et al. 2014], and CODEX [Jiang et al. 2015]) with the ability to incorporate results from additional programs such asXHMM [Fromer and Purcell 2014]. In addition, our pipeline: 1) Generates read counts only once, either from BAM or CRAM; 2) Runs the three methods in parallel; 3) Merges calls by a user-defined overlap percentage and a size threshold; 4) Provides annotation such as gene names in the regions and call frequencies.

Materials and methods

Sample selection:

- 1,633 BHCMG samples with WES data.

Sequencer and reagents:

- Exome Capture: Agilent SureSelect HumanAllExonV4 or V5 plus clinical content.
- Illumina HiSeq2500 platform (Majority).
- TruSeq Rapid SBS-HS 100 bp Paired Ends (Majority).

Sequencing data processing:

- BWA mem 0.7.8 alignment, local alignment and base call quality score recalibration with GATK 3.1-1.

Exome CNV calling programs:

- ExomeDepth, XHMM, CANOES and CODEX.

Results

The ECO algorithm:

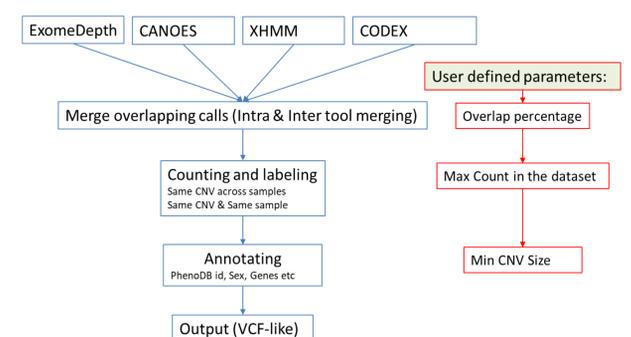
First pull the results across individuals and programs all together, then separate them by chromosomes and sort the calls by decreasing lengths, then for each chromosome, beginning with the largest CNV call (CNV_1), we assign the same unique ID if any of the smaller CNVs (CNV_2) overlap with CNV_1 and $\text{length}(\text{CNV}_2)/\text{length}(\text{CNV}_1) \geq \text{overlap percentage threshold}$. The process continues until each call has been assigned with an ID.

The pipeline: Python job control scripts adapted from UW-GAC QCpipeline (<https://github.com/UW-GAC/QCpipeline>)

CNV call summary:

	ExomeDepth	XHMM	CANOES	CODEX
Total	188,703	38,952	10,607	259,146
Median	85	22	6	43
Range	30 ~ 1113	5 ~ 276	1 ~ 97	16 ~ 8650
#calledSamples	1633	1563	1603	1633

ECO flow chart:



An example output:

```

## Run on 27/05/17 02:12
## combine cnv results
## Filters: SET_COUNT = 15 SET_OVERLAP_PCT = 0.8 SET_CNVS_SIZE = 100
## count_in_data: times of the same cnv (same cnv_ID) called in the dataset from different individuals
## count_in_sample: times of same cnv (same cnv_ID) called from the same individual
## size: cnv sizes in bp, base pair
## cnv_ID: IDs assigned to each unique cnv, with format chromosome_num, where same cnv_ID indicates the same cnv using current filters
## Info: other information from each program if that cnv was called by that program
sm_tag chr start end count_in count_in_size cnv_ID PhencSex Project #ofGenes firstFiveG Info
10121-110 21 85617954 1.46E+08 1 1 60661764 21_2162 BH692 F M_Valle_I . canoes_info:type:MI
10121-110 8 85785261 1.29E+08 1 1 43377424 8_6045 BH692 F M_Valle_I 248 LOC10192 XHMM-type:MID_BP
10121-110 21 35401736 77912412 1 1 42510676 21_2161 BH692 F M_Valle_I 198 KCNE2,LO canoes_info:type:MI
33654-112 4 91341197 1.27E+08 1 1 35888621 4_8126 BH673 F M_Valle_I 174 SNORA24,canoes_info:type:MI
  
```

Summary

- We proposed a pipeline, ECO, for integrating CNV results from different programs, which allows users to merge with overlap percentage and to filter results based on the CNV size and frequency.
- Read counts generated can be used by multiple programs, no need to go back to BAM/CRAM files.
- This framework can be extended to include results from other programs such as and HMZDeFinder [Gambin et al. 2017] and EXCAVATOR [Magi et al. 2013] and whole genome sequencing.
- Future work includes further refinement of calls from each individual program and cross validation across programs.